

## **Procedure for Blood Analysis**

**1.0 Purpose** - This procedure specifies the methods for performing blood analysis in forensic casework.

**2.0 Scope** - This procedure applies to those Forensic Scientists who have been released, to the extent of their training, to do blood analysis in forensic casework.

**3.0 Definitions** - N/A

### **4.0 Equipment, Materials and Reagents**

- Ethyl alcohol, 200 proof, anhydrous, 99.5+%
- Phenolphthalein (see QC procedure)
- 3 % hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) (see QC procedure)
- Whatman 55 mm filter papers/swabs
- Known blood stain
- Sterile disposable scissors or scalpel blade
- RSID kits which contain the test cards and universal buffer
- 1.5 mL centrifuge tube

### **5.0 Procedure**

#### **5.1 Kastle-Meyer Test**

**5.1.1** The Kastle-Meyer Test (KM) shall be performed on stained areas that appear reddish brown in color. If multiple stains are present, the Forensic Scientist shall determine which stains shall be tested based upon his/her training and experience. If blood examination is requested and the item does not appear to have any stains, the entire item shall be examined. If this general rubbing is negative, pictures are not required to be taken and notes shall reflect that a general rubbing was made and the item was negative. If the general rubbing is positive, but no visible stain can be located, the notes in FA shall reflect that a general rubbing was positive but no stains were isolated. A photograph shall be taken of the item to show the general area that tested positive. If latent or touch DNA has been requested or ridge detail is noted, swabbing the item would compromise possible prints or DNA. In this situation, only a visual examination shall be done on this item.

**5.1.2** Each stain that is tested shall be numbered and a photograph shall be taken to show the area(s) that were examined. The only exception is stained swabs. The results of each area tested shall be noted in FA as positive or negative.

**5.1.3** Rub the area to be tested with a folded piece of filter paper or a clean cotton swab.

**5.1.4** Add the following reagents onto the filter paper or swab in order:

- 1-2 drops of ethanol (cover rubbed area).
- 1-2 drops of phenolphthalein (cover rubbed area).
- 1-2 drops of 3 % H<sub>2</sub>O<sub>2</sub> (cover rubbed area).

**5.1.5** If a suspected area tests negative for KM with a rubbing, further testing shall be done. Take a small cutting with a sterile disposable scissors or scalpel blade and place on a filter paper. Add the chemicals directly to the sample.

### **5.1.6 Results**

**5.1.6.1** A positive reaction is indicated by the development of a pink color within 5 seconds of adding the  $H_2O_2$ .

**5.1.6.2** A negative result is indicated if no color change occurs within 5 seconds after the addition of  $H_2O_2$ . In all situations, a color change will eventually occur after the addition of  $H_2O_2$ . This color change does not indicate a positive reaction; therefore, any color changes that appear after 5 seconds should not be interpreted as a positive result.

**5.1.6.3** An inconclusive result is indicated if a reaction occurs before the addition of the  $H_2O_2$  or if color transfers from the item onto the filter paper or swab that interferes with the ability to note a color change. If this occurs, a RSID blood test shall be performed if enough material is present to perform this test.

## **5.2 RSID-Blood Testing**

### **5.2.1 Procedure**

**5.2.1.1** Cut a small sample, approximately 0.5 cm<sup>2</sup> (depending on the concentration of the stain), from the evidence sample using sterile disposable scissors or a sterile scalpel blade and place the cutting into a 1.5 mL centrifuge tube.

**5.2.1.2** Add a minimum of 150 µL, up to 1 mL, of RSID universal buffer to each sample and mix well. (The amount of buffer added will depend on the sample size; buffer should cover the sample completely.)

**5.2.1.3** Allow the sample to extract for a minimum of 15 minutes. For weaker or older samples, Forensic Scientists should use a larger quantity of material and/or an extended extraction time to include overnight.

**5.2.1.4** After completing the extraction process, pipette 100 µL of the extracted sample into the sample well on the RSID card.

### **5.2.2 Results**

**5.2.2.1** A positive reaction will have two lines appear in the test window. One line will appear in the area marked "C" for control and one line will appear in the area marked "T" for test. A positive result may be recorded as soon as both of these lines appear, but after no longer than 10 minutes. The lines must be reddish in color.

**5.2.2.2** If a line does not appear in the "T" area within ten minutes, then the test is considered negative. A line must appear at the area marked "C" to ensure that the test is working properly.

**5.2.2.3** If no line appears at the area marked "C," the test shall be repeated. If no line is seen in the "C" window in the repeated test, the test is considered inconclusive. If this occurs, the Body Fluid Technical Leader shall be notified as soon as possible. Refer to Forensic Biology Section Administrative Policy and Procedure.

**5.2.3 Requirements for testing** - The RSID Blood Test shall be performed if human blood identification is requested specifically, species origin is in question, or the Kastle-Meyer test is inconclusive, as provided in **5.1.6.3**. The only exception would be if there is a limited amount of sample and DNA testing is requested.

**5.3 Reporting guidelines** – The results statements shall reflect only the work that is performed. Portions of the statements may be omitted to address the testing actually performed. This interpretation may include or build upon one (1) or more of the following responses depending on the circumstances of the case and the nature of the examination.

**5.3.1** This phrase shall be used when a negative result is indicated for a Kastle-Meyer Test.

Examination of a sample(s)/general rubbing(s) taken from \_\_\_\_\_ (Item(s) \_\_\_\_), using the Kastle-Meyer Test, failed to reveal chemical indications for the presence of blood.

**5.3.2** This phrase shall be used when a positive result is indicated for a Kastle-Meyer Test.

Examination of a sample(s)/general rubbing(s) taken from \_\_\_\_\_ (Item(s) \_\_\_\_), using the Kastle-Meyer Test, gave chemical indications for the presence of blood.

**5.3.3** This phrase shall be used when an inconclusive result is indicated for a Kastle-Meyer Test.

Examination of a sample(s)/general rubbing(s) taken from \_\_\_\_\_ (Item(s) \_\_\_\_), using the Kastle-Meyer test, failed to reveal conclusive results for chemical indications for the presence of blood because of possible interference of the substrate.

**5.3.4** This phrase shall be used if no confirmatory blood testing was done.

No confirmatory blood testing was performed.

**5.3.5** This phrase shall be used if no confirmatory blood testing was done due to limited sample.

Due to the limited quantity of sample, no confirmatory blood testing was done.

**5.3.6** This phrase shall be used when the confirmatory RSID blood test yields a negative result:

Further examination of the sample(s) taken from \_\_\_\_\_ (Item(s) \_\_\_\_), using the RSID Blood test, failed to reveal the presence of human blood.

**5.3.7** This phrase shall be used when the confirmatory RSID Blood Test yields a positive result:

Further examination of the sample(s) taken from \_\_\_\_\_ (Item(s) \_\_\_\_), using the RSID Blood test, gave conclusive results for the presence of human blood.

## **5.4 Controls**

**5.4.1 Kastle-Meyer Controls:** A known blood stain shall be used as a positive control and a clean filter paper or swab shall be used for a negative control. These controls shall be tested prior to analysis once each day this test is performed for each lot used and the results shall be recorded as positive or negative in the case notes in FA for each case worked that day. The controls must react appropriately.

**5.4.2 RSID-Blood Controls:** A positive control (applicable body fluid standard), and a negative control (100 µL of universal buffer) shall be run with every case or every batch of cases and the results will be recorded in the case notes as a positive or negative. If a reddish line is seen in the negative control “T” area, the test shall be rerun. If a reddish line appears again in the negative control “T” area, the test shall be considered inconclusive. If this occurs, the Technical Leader shall be notified immediately.

**6.0 Limitations** – Limitations include, but are not limited to, the following: The Kastle-Meyer test is a presumptive test for the presence of blood. There are some substances that will cross-react giving a false positive. If a stain yields a positive result and the analyst suspects the result may be a false positive based on training and experience, the area may be rubbed with filter paper and placed in an incubator at 100 °C for 30 minutes. The filter paper shall then be tested as indicated in **5.1.4**. The result of this test shall be recorded in the FA worksheet.

**7.0 Safety** - See Safety documents.

## **8.0 References**

Forensic Biology Section Body Fluid training documents

Forensic Biology Section Procedure for Aseptic Technique and Contamination Control

Forensic Biology Section Procedure for Calibration and Maintenance

**9.0 Records** - N/A

**10.0 Attachments** - N/A

Revision History		
Effective Date	Version Number	Reason
10/26/2012	1	Original Document - Combined Procedure for Kastle-Meyer test and RSID-blood portion of the RSID Procedure; added reporting guidelines and allowed for changes to be made by the Forensic Scientist to address the testing actually performed; 5.2.3 added requirement to perform RSID-blood testing.
02/01/2013	2	5.1.1 – added requirement for testing item with multiple stains present; 5.4.1 – added requirement for QC testing each lot used
02/15/2013	3	5.3.4, 5.3.5 – changed “no further confirmatory” to “no confirmatory” 6.0 – clarified wording